## Supplementary Information for

## MRBLE-pep measurements reveal accurate binding affinities for B56, a PP2A regulatory subunit.

Jamin B. Hein<sup>1,2,3</sup>, Martha S. Cyert<sup>1</sup>, Polly M. Fordyce<sup>2,4,5,6</sup>

<sup>1</sup> Department of Biology, Stanford University, Stanford, CA 94305

Blegdamsvej 3b, 2200 Copenhagen, Denmark

<sup>4</sup> Department of Genetics, Stanford University, Stanford, CA 94305

<sup>5</sup> ChEM-H Institute, Stanford University, Stanford, CA 94305

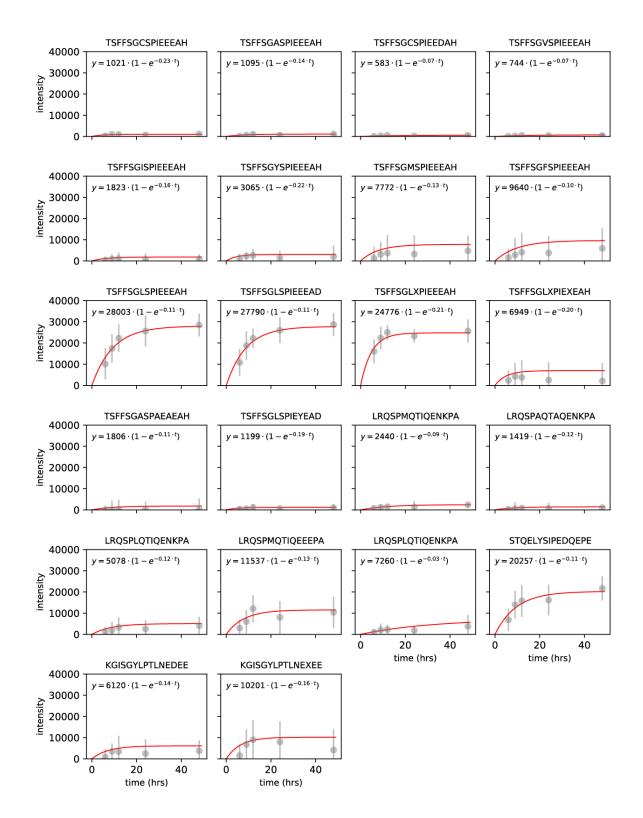
<sup>6</sup> Chan Zuckerberg Biohub, San Francisco, CA 94110

<sup>&</sup>lt;sup>2</sup> Department of Bioengineering, Stanford University, Stanford, CA 94305

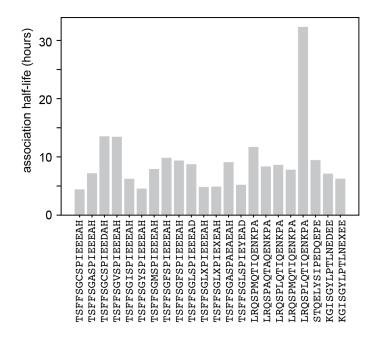
<sup>&</sup>lt;sup>3</sup> The Novo Nordisk Foundation Center for Protein Research, Faculty of Health and Medical Sciences, University of Copenhagen,

Code	Sequence	Mean (Kd)	SE (Kd)	Mean ( $\Delta\Delta$ G)	SE ( $\Delta\Delta$ G)	Library
1	TSFFSGCSPIEEEAH	71073.44	10306.54	2.82	0.23	1
2	TSFFSGASPIEEEAH	104654.44	19522.14	3.04	0.19	1
3	TSFFSGCSPIEEDAH	158673.79	20642.92	3.30	0.28	1
4	TSFFSGVSPIEEEAH	344022.43	57937.01	3.76	0.28	1
5	TSFFSGISPIEEEAH	77163.64	3446.67	2.88	0.31	1
6	TSFFSGYSPIEEEAH	40576.78	6039.53	2.49	0.19	1
7	TSFFSGMSPIEEEAH	39828.93	15393.48	2.39	0.12	1
8	TSFFSGFSPIEEEAH	21125.87	3881.26	2.10	0.19	1
9	TSFFSGLSPIEEEAH	824.49	299.18	0.07	0.02	1
10	TSFFSGLSPIEEEAD	719.33	248.05	0.00	0.00	1
11	TSFFSGLXPIEEEAH	430.93	157.18	-0.32	0.03	1
12	TSFFSGLXPIEXEAH	25598.31	4459.53	2.21	0.17	1
13	TSFFSGASPAEAEAH	283395.36	38820.16	3.64	0.35	1
14	TSFFSGLSPIEYEAD	92865.40	8706.23	2.99	0.25	1
15	LRQSPMQTIQENKPA	44031.44	11871.24	2.51	0.29	1
16	LRQSPAQTAQENKPA	77891.82	18278.86	2.86	0.28	1
17	LRQSPLQTIQENKPA	27692.73	3245.71	2.27	0.29	1
18	LRQSPMQTIQEEEPA	12766.19	4588.22	1.73	0.12	1
19	LRQSPLQTIQENKPA	27809.86	4479.58	2.27	0.29	1
20	STQELYSIPEDQEPE	1893.03	632.36	0.59	0.06	1
21	KGISGYLPTLNEDEE	43087.35	10136.94	2.50	0.16	1
22	KGISGYLPTLNEXEE	13148.79	4776.67	1.71	0.05	1
1	DFTRLQDIPEETESR	755.78	313.65	0.43	0.04	2
2	DFTRMQDIPEETEXR	11812.68	2771.72	2.14	0.09	2
3	DFTRAQDAPAETESR	39734.00	12185.37	2.83	0.21	2
4	DFTRMQDIPEETESR	10706.87	2184.70	2.09	0.12	2
5	NKRLSTIDEXGSILS	311.00	76.28	0.18	0.06	2
6	NKRLSTIDEEGSILS	161.85	47.09	-0.43	0.04	2
7	NKRASTADASGSILS	25288.43	6720.82	2.58	0.12	2
8	NKRLSTIDESGSILS	3947.35	139.40	1.52	0.20	2
9	LRQSPLQTIGEEEPA	1216.59	635.59	0.67	0.12	2
10	LRQXPLQTIQEEEPA	346.25	119.03	0.00	0.00	2
11	LRQSPLKTIKEEEPA	54.63	18.22	-1.08	0.08	2
12	LRQSPLQTIQEEEPA	124.51	35.18	-0.58	0.06	2
13	LRQSPMQTIQEEEPA	6576.38	773.96	1.82	0.15	2
14	LRQSPLQTIQENKPA	2501.93	753.05	1.20	0.08	2
15	LRQSPAQTAQENKPA	32042.00	9690.55	2.71	0.16	2
16	LRQSPMQTIQENKPA	9325.89	886.41	2.03	0.18	2
17	TSFFSGASPAEAEAH	31616.29	8755.70	2.71	0.18	2
18	TSFFSGLXPIEEEAH	119.18	30.79	-0.60	0.07	2
19	TSFFSGLSPIEEEAH	128.32	41.73	-0.58	0.02	2
20	TSFFSGFSPIEEEAH	581.48	224.80	0.30	0.06	2
21	TSFFSGISPIEEEAH	12282.41	2181.89	2.18	0.15	2
22	TSFFSGVSPIEEEAH	21912.73	5620.81	2.50	0.11	2
23	TSFFSGCSPIEEDAH	21088.21	5071.72	2.48	0.12	2
24	TSFFSGCSPIEEEAH	9551.44	1878.54	2.02	0.12	2

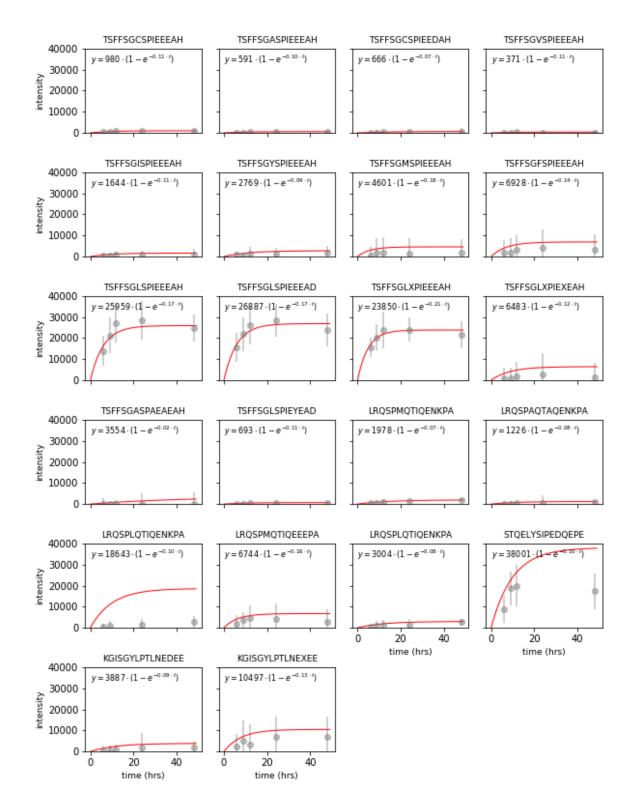
**Table S1**: Mean  $K_d$  (nM) and mean  $\Delta\Delta G$  (kcal/mol) for peptides calculated from triplicate measurements for both libraries.



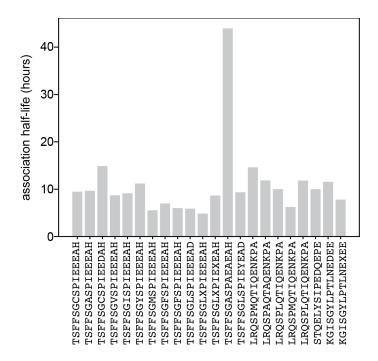
**Fig. S1**. Measurement of time required to reach binding equilibrium for all peptides in MRBLE-pep library 1. Grey markers indicate median bead intensity at a given time point, error bars indicate the standard deviation, and the red line indicates a fit to a kinetic binding curve (equation and fit parameters given at top).



**Fig. S2**. Association half-life for all peptides from MRBLE-pep library 1 (determined from kinetic binding fit parameters).



**Fig. S3**. Measurement of time required to reach binding equilibrium for all peptides in MRBLE-pep library 1. Grey markers indicate median bead intensity at a given time point, error bars indicate the standard deviation, and the red line indicates a fit to a kinetic binding curve (equation and fit parameters given at top).



**Fig. S4**. Association half-life for all peptides from MRBLE-pep library 1 (determined from kinetic binding fit parameters).





anti- mouse secondary Antibody only

	anti-B56
100 mm	

**Fig. S5**. Western Blot with the unbound fractions of a MRBLE-pep concentration experiment. The unbound fraction from each concentration of a MRBLE-pep concentration experiment was probed for unbound B56 protein.

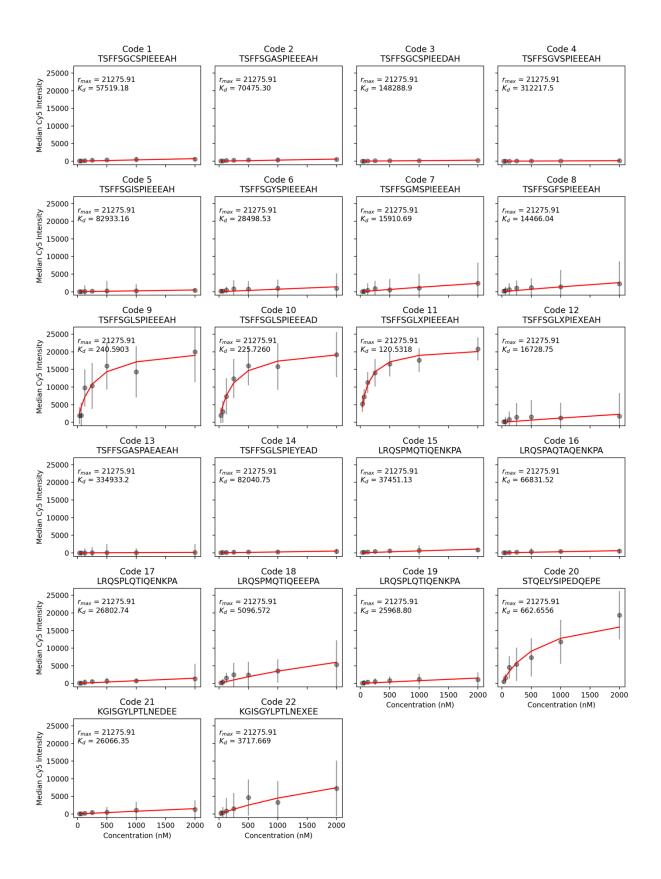
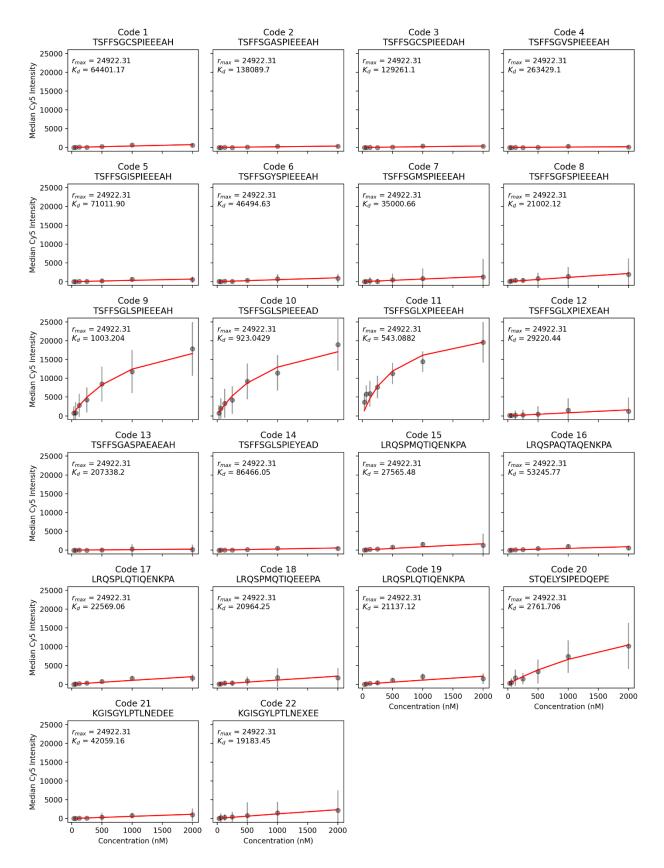


Fig. S6. Concentration-dependent binding data (grey markers, median intensity and standard deviation over all beads) and associated Langmuir isotherm fits (red lines) for MRBLE-pep library 1, replicate #1.



**Fig. S7**. Concentration-dependent binding data (grey markers, median intensity and standard deviation over all beads) and associated Langmuir isotherm fits (red lines) for MRBLE-pep library 1, replicate #2.

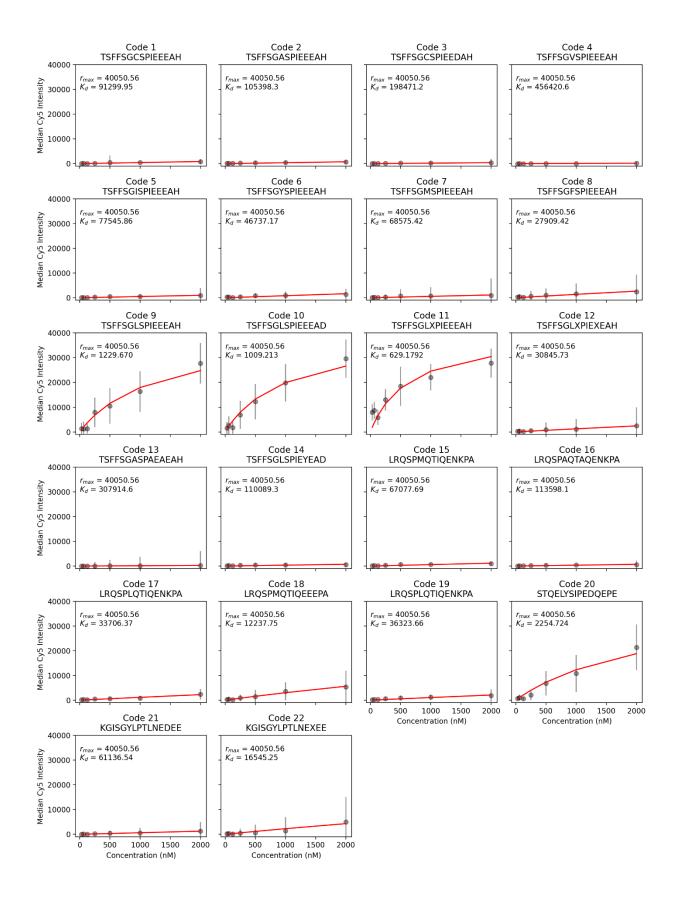
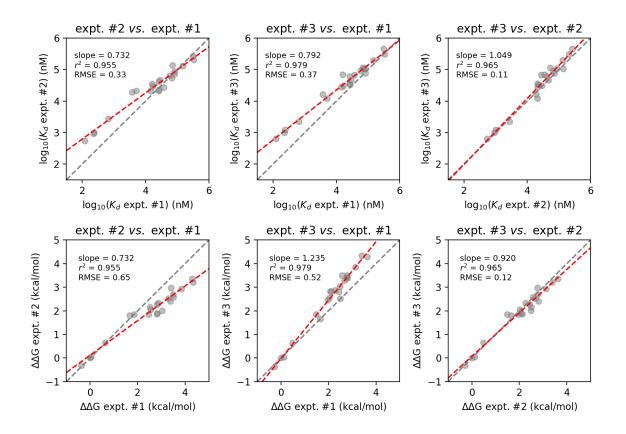
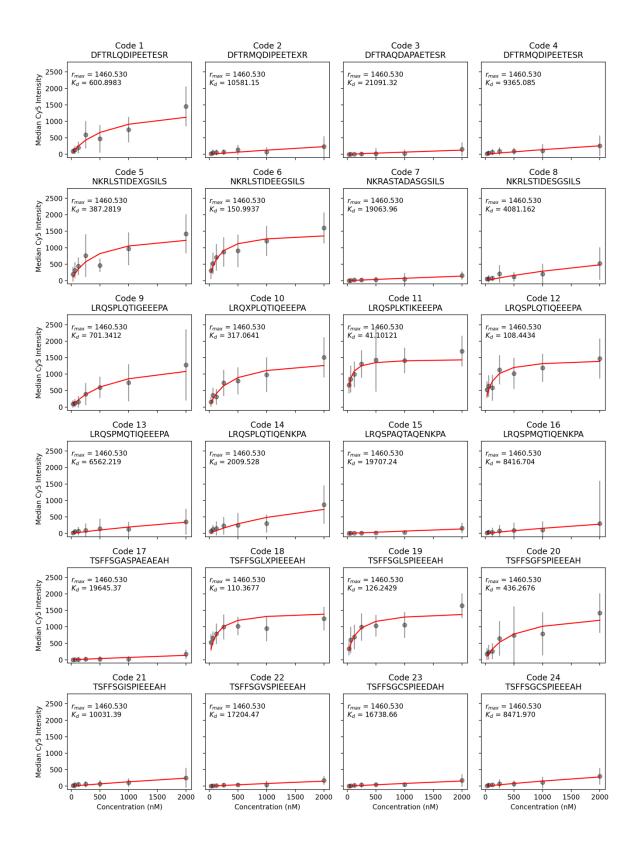


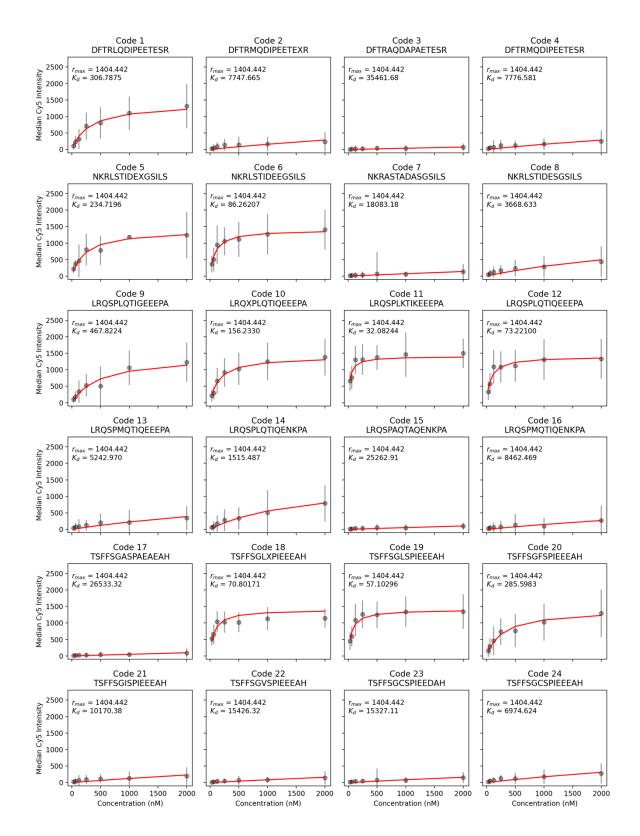
Fig. S8. Concentration-dependent binding data (grey markers, median intensity and standard deviation over all beads) and associated Langmuir isotherm fits (red lines) for MRBLE-pep library 1, replicate #3.



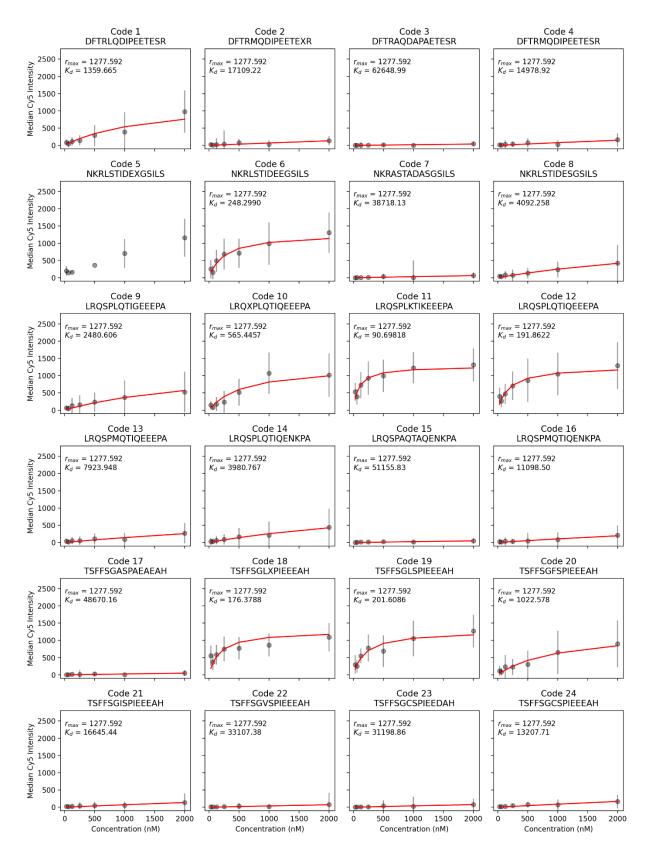
**Fig. S9**. Comparison of measured affinities between technical replicates for MRBLE-pep library 1. **(A)** Comparison of  $K_{ds}$  for each B56-peptide interaction (returned values from global Langmuir isotherm fits). Black dashed line indicates the 1:1 identity line; red dashed line indicates a linear regression to  $\log_{10^-}$ transformed  $K_{d}$  values. **(B)** Comparison of  $\Delta\Delta$ Gs for each B56-peptide interactions (calculated relative to a Kif4A 'reference' peptide sequence of TSFFSGLSPIEEEAD). Black dashed line indicates the 1:1 identity line; red dashed line indicates a linear regression.



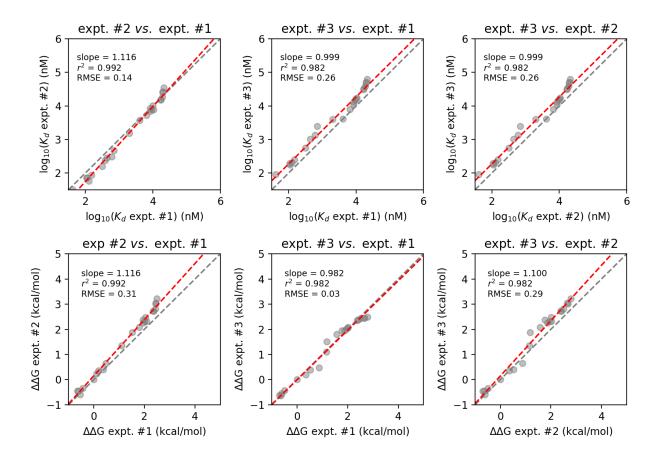
**Fig. S10**. Concentration-dependent binding data (grey markers, median intensity and standard deviation over all beads) and associated Langmuir isotherm fits (red lines) for MRBLE-pep library 2, replicate #1.



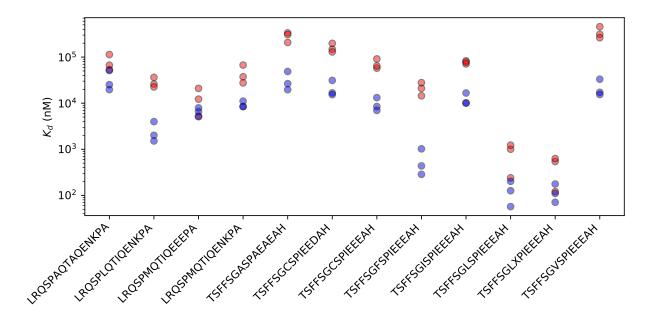
**Fig. S11**. Concentration-dependent binding data (grey markers, median intensity and standard deviation over all beads) and associated Langmuir isotherm fits (red lines) for MRBLE-pep library 2, replicate #2.



**Fig. S12**. Concentration-dependent binding data (grey markers, median intensity and standard deviation over all beads) and associated Langmuir isotherm fits (red lines) for MRBLE-pep library 1, replicate #3.



**Fig. S13**. Comparison of measured affinities between technical replicates for MRBLE-pep library 2. **(A)** Comparison of  $K_ds$  for each B56-peptide interaction (returned values from global Langmuir isotherm fits). Black dashed line indicates the 1:1 identity line; red dashed line indicates a linear regression to  $\log_{10^-}$ transformed  $K_d$  values. **(B)** Comparison of  $\Delta\Delta$ Gs for each B56-peptide interactions (calculated relative to a Kif4A 'reference' peptide sequence of TSFFSGLSPIEEEAD). Black dashed line indicates the 1:1 identity line; red dashed line indicates a linear regression.



**Fig. S14**. Measured  $K_{ds}$  for 12 peptides containing Kif4A-like or FoxO3-like motifs across 3 technical replicates each of MRBLE-pep library 1 (red markers) and library 2 (blue markers).